

## Original article

**Gut microbiota involved in sleep quality mediated by the gut-brain axis**Haasbroek Kyle<sup>1)</sup>, Mari Ogura<sup>1,2)</sup>, Masayuki Yagi<sup>1)</sup>, Yoshikazu Yonei<sup>1)</sup>

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**Abstract**

Through the mediation of the gut-brain axis, sleep quality has a bidirectional effect on the gut microbiota. Using fecal T-RFLP flora analysis, we have found that improving sleep quality leads to changes in the gut microbiota. In particular, we observed an increase in short-chain fatty acid (SCFA)-producing bacteria such as *Bacteroidota*, *Oscillospiraceae*, and *Lachnospiraceae*. The microbiota of persons with poor “sleep quality” is similar to the pattern of dementia patients, and may approach the pattern of healthy people as “sleep quality” improves. In this report, we picked up major bacteria that are expected to be related to sleep quality and reviewed the literature. It seems important to avoid dysbiosis and maintain the dominance of SCFA-producing bacteria in order to maintain good “sleep quality” and hopefully prevent the progression of dementia.

**KEY WORDS:** microbiota, sleep quality, dementia, short-chain fatty acids

**Introduction**

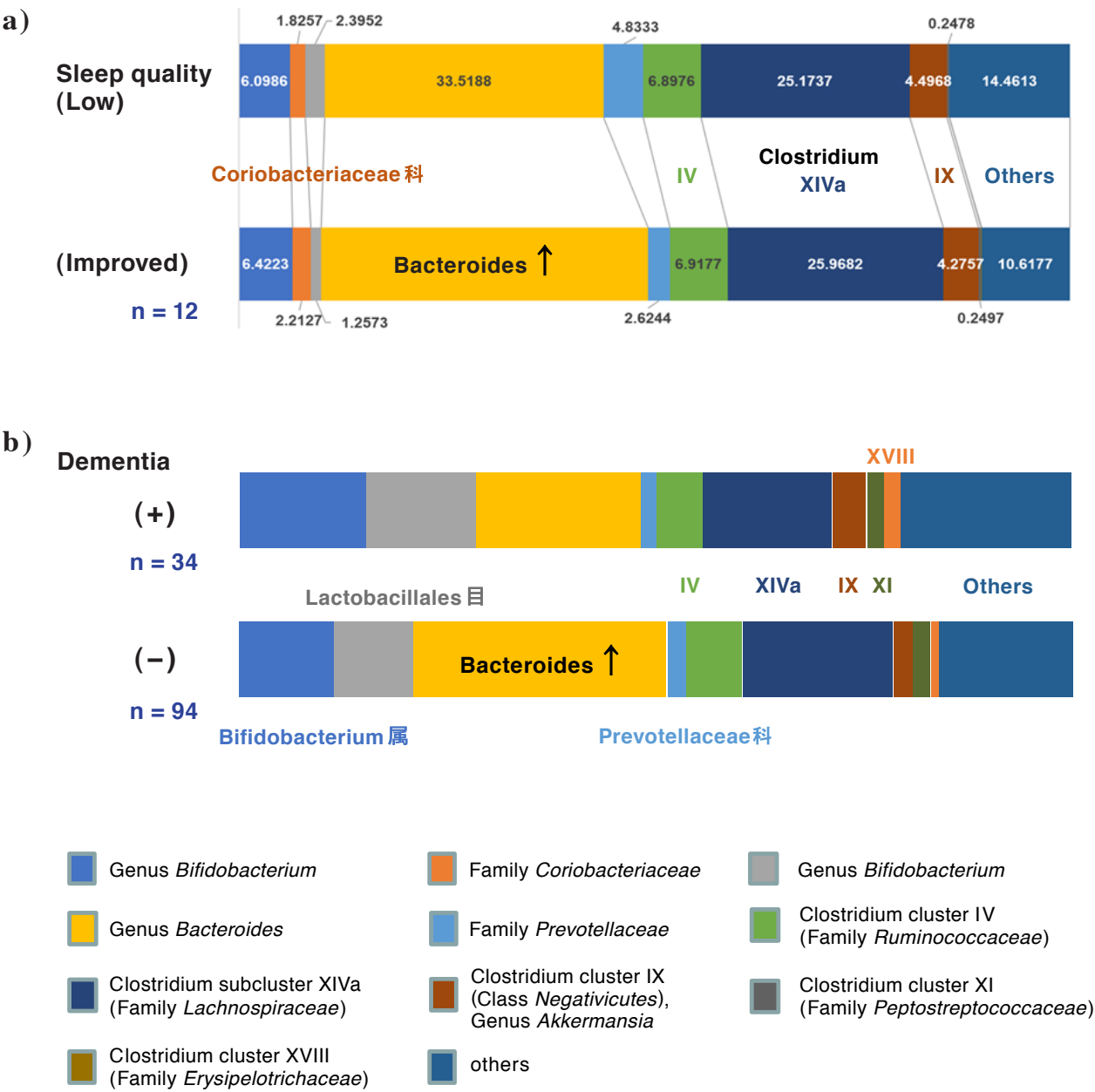
Six clinical trials have shown that various physical functions improve with improved sleep quality<sup>1-6)</sup>. For PSQI-J, sleep quality improved significantly in all six trials, sleep onset time improved significantly in five out of six trials, sleep duration improved significantly in four out of six trials, and difficulty waking during the day improved significantly in three out of six trials. The overall assessment PSQIG improved significantly in all six trials. For the third trial, a crossover study was conducted with a control group, but the improvement in the subject groups was similar<sup>3)</sup>. Similar results were obtained in all six trials, showing that the improvement in subjective symptoms was highly reproducible and that the use of the test bedding improved “sleep quality.”

In the fifth trial, the relationship between sleep quality, intestinal flora, and amyloid beta (A $\beta$ ) clearance index

(A $\beta$ 40/42) was examined<sup>5)</sup>. As a result, an increase in short-chain fatty acid (SCFA)-producing bacteria was observed in the gut microbiota, indicating the influence of the gut-brain axis (**Fig.1-a**). The A $\beta$ 40/42 of the subjects was significantly higher than that of healthy subjects, suggesting the possibility that A $\beta$  clearance may be reduced in subjects with poor “sleep quality.”

Saji et al. compared the gut microbiota of dementia patients and healthy subjects and reported that the healthy subjects had more short-chain fatty acid-producing bacteria (**Fig.1-b**)<sup>7)</sup>. This is a very interesting example of brain-gut coupling.

In this study, we focused on gut bacteria related to dementia and sleep quality in these clinical trials<sup>5,7)</sup> and summarized recent findings.



**Fig. 1. Comparison with other research findings.**

**a)** The gut microbiota of subjects with reduced "sleep quality". The microbiota composition shifted as "sleep quality" improved.  
**b)** The difference in microbiota composition between dementia patients and healthy subjects. Figure (a) quoted from Reference 5 and Figure (b) quoted and modified from Reference 7.

## Methods

### Gut microbiota

It is known that the human intestinal lumen is home to approximately 1,000 species and over 100 trillion intestinal bacteria that form a stable community (gut microbiota,) while forming a symbiotic and antagonistic relationship with each other. The bacteria that make up the gut microbiota interact with the host directly or through their metabolites, affecting our metabolic functions and immune system<sup>8-11</sup>, and furthermore, bidirectionally affecting the central nervous system via the gut-brain axis<sup>12,13</sup>.

Next-generation DNA sequencer technology is used to analyze the gut microbiota, and 16S rRNA amplicon analysis and shotgun whole metagenomic analysis are being performed. The 16S ribosomal RNA gene is said to be an essential gene for bacteria, and bacteria are classified based on the similarity of the base sequence. The unit obtained when the base sequence of the bacterial 16S ribosomal RNA gene is classified on a computer based on its similarity (generally 96-97%) is called an OTU (operational taxonomic unit). The number of OTUs represents the number of bacterial species that compose the bacterial flora, and the number of reads belonging to the same OTU is thought to represent the relative abundance of that species.

In this study, the major taxonomic groups of the human intestinal flora (*Bifidobacterium*, *Bacteroides*, etc.) were analyzed as OTUs. Bacteria related to those previously reported<sup>5,7</sup> were selected and a literature review was added.

## Results

### Changes in the intestinal flora

In fecal T-RFLP flora analysis, the proportion of *Bacteroides* genus bacteria was  $33.5 \pm 8.2\%$  before use and  $39.4 \pm 7.3\%$  after 8 weeks of use, showing a significant increase after 8 weeks compared to before use ( $p < 0.001$ , Fig. 1-a)<sup>5</sup>. In addition to *Bacteroides*, an increase in SCFA-producing bacteria such as *Oscillospiraceae* and *Lachnospiraceae* was also observed. Analysis of other bacteria seems necessary. The importance of major bacteria to health is summarized below.

### Genus *Caproicibacter*

Belonging to the family *Oscillospiraceae*, the genus *Caproicibacter* is comprised of a single known species, *Caproicibacter fermentans*, first described in 2020<sup>14</sup> after isolation from a gas bioreactor in Germany. *C. fermentans* is an obligate anaerobic, spore-forming bacterium with a gram positive-like cell wall structure despite staining gram-negative. Little is known of *C. fermentans* ecological role in the gut microbiome or its effects on human health.

*C. fermentans* is capable of metabolizing a variety of saccharides, including glucose, fructose, and sucrose. As metabolic byproducts, the bacterium produces SCFAs acetate, butyrate, and lactate, as well as the medium chain fatty acid caproate, ethanol, CO<sub>2</sub>, and H<sub>2</sub>.

The physiological significance of this bacterium is unclear.

### Genus *Lachnospiraceae incertae sedis*

*Lachnospiraceae incertae sedis*, as the name states, is a taxon which contains bacteria of uncertain phylogeny and lacking description within the family *Lachnospiraceae*. The taxon contains an abundance of candidate species which have been genetically identified<sup>15</sup> but have not reportedly been cultured and lack a detailed metabolic description. Nevertheless, modern genomic techniques have allowed the detection of *L. incertae sedis* in a variety of contexts.

The taxon is reported to be abundant in those in better cardiovascular health<sup>16</sup> and in those without Crohn's disease<sup>17</sup>. Loss of *L. incertae sedis* may also serve as a biomarker for hepatocellular carcinoma, as it is significantly reduced in mouse models of the disease<sup>18</sup>. On the other hand, *L. incertae sedis* is enriched in non-alcoholic fatty liver disease<sup>19</sup>, type 2 diabetes<sup>20</sup>, alopecia areata<sup>21</sup>, and is associated with major depressive disorder<sup>22,23</sup>. In infants, the taxon is more abundant with eczema<sup>24</sup> and correlated with lower mental development index scores<sup>25</sup>. As uncertain as its taxonomy, the observed relationship with human health is also unclear, revealing potentially both beneficial and pathogenic relationships.

The relationship with human health is still unclear, and since there is a possibility of both beneficial and pathogenic relationships, it is difficult to evaluate the physiological significance. However, these bacteria produce butyric acid and are known to be common in long-lived people<sup>26</sup>, thus an increase in this taxon may be considered a positive effect. It is likely that the contradictory positive and negative health correlations with this taxon may be due to disparate effects of individual member species which are still awaiting an agreed upon taxonomic classification.

### Genus *Aminipila*

*Aminipila* is a recently discovered genus, first described in 2018<sup>27</sup> when the type species *Aminipila butyrica* was isolated from a cattle waste bioreactor in Japan. Subsequently, two more species, *Aminipila terrae*<sup>28</sup> and *Aminipila luticellarii*<sup>29</sup>, have been named. Information regarding the role of *Aminipila* in the gut microbiome remains sparse. *A. butyrica* is a strictly anaerobic gram-positive microbe. The species is asaccharolytic, relying on the fermentation of amino acids arginine, lysine, and serine as a growth substrate, producing the SCFAs acetate and butyrate as well as hydrogen sulfide as metabolic byproducts.

In humans, *A. butyrica* has been detected in the subgingival microbiome where its abundance may serve as a biomarker for rheumatoid arthritis<sup>30</sup>. However, experimental evidence of its role in the gut is lacking. *A. butyrica* has also been identified in the gut of sheep, where it promotes resistance to parasitic infection by helminth<sup>31</sup>.

This bacterium may be beneficial in that it increases SCFA production and provides resistance to parasitic infections.

### Genus *Butyricimonas*

*Butyricimonas* are a genus of obligate anaerobic gram-negative bacteria named for producing large amounts of butyric and isobutyric acid<sup>32</sup>, and first isolated from rat fecal samples<sup>33</sup>. Metabolizing glucose, the major metabolic SCFA

endproducts are butyrate, isobutyrate, and isovalerate, with smaller amounts of acetate, propionate, and succinate<sup>34</sup>). The genus contains species such as *B. faecalis*, *B. faecihominis*, *B. paravirosa*, *B. synergistica*, and *B. virosa*.

*Butyricimonas* is characterized by beneficial bacteria that perform an important function in SCFA production and the maintenance of intestinal homeostasis. As a significant butyrate producer, *Butyricimonas* contributes to maintaining intestinal barrier function and reduces inflammation. In mouse models of diabetes<sup>35</sup>) and metabolic syndrome<sup>36</sup>), ameliorating treatment was associated with enrichment of *Butyricimonas* and improvement of intestinal dysbiosis (e.g., increased SCFAs and reduced inflammation, intestinal barrier function, serum LPS). Furthermore, intake of *B. virosa* has demonstrated a protective effect against high fat diet induced diabetes in mice<sup>37</sup>). Additionally, in menopausal women increased intestinal production of SCFA butyrate was associated with increased muscle mass and abundance of *B. virosa*<sup>38</sup>).

*Butyricimonas* abundance is associated with reduced BMI, and it is reduced by diets high in animal protein, saturated fats, and simple sugars<sup>39</sup>). In the successful treatment of chronic constipation with fecal microbiota transplant, *Butyricimonas* was significantly enriched in abundance after treatment<sup>40</sup>).

*Butyricimonas* may also influence immune regulation and inflammation, as it is significantly reduced in multiple sclerosis patients, both with and without treatment, compared to healthy controls and its abundance is negatively correlated with proinflammatory gene expression characteristic of MS<sup>41</sup>). *Butyricimonas* is similarly reduced in patients with histamine intolerance<sup>42</sup>), associated with an inflammatory intestinal reaction in response to intake of foods containing histamine. *Butyricimonas* is also significantly reduced in Cystic Fibrosis, and its abundance negatively correlated with gene expression associated with colorectal cancer<sup>43</sup>).

An increase in these butyric acid-producing bacteria can be seen as a favorable change.

### Genus *Hydrogenoanaerobacterium*

The genus *Hydrogenoanaerobacterium* contains a single known species, *H. saccharovorans*. The bacterium is an obligate anaerobic gram-negative microbe that was initially isolated from a hydrogen producing bioreactor in China<sup>44</sup>). As the name suggests, *H. saccharovorans* is supported by metabolizing a wide range of saccharides. The main endproducts of its glucose metabolism are ethanol, acetate, hydrogen, and carbon dioxide. *Hydrogenoanaerobacterium* has been detected as a member of the human gut microbiome in several studies, although the implications of *H. saccharovorans* for human health remains unclear, with most of the current data extracted from animal experimentation using mice.

In a mouse model of high fat diet induced obesity, oolong tea extract reduced weight gain and ameliorated lipid metabolism; *Hydrogenoanaerobacterium* was significantly enriched in the treatment group<sup>45</sup>). In mouse model of alcoholic liver, ameliorating treatment with a fungal extract resulted in increased abundance of *Hydrogenoanaerobacterium*, among other taxa<sup>46</sup>). Conversely, it has also been reported that *Hydrogenoanaerobacterium* is elevated in high fat diet-fed mice, and was reduced by treatment with resveratrol via repression of mTOR Complex 2<sup>47</sup>).

With human data, the associations are notably more negative. *Hydrogenoanaerobacterium* abundance was observed to be significantly increased in sporadic Parkinson's Disorder patients<sup>48</sup>) and in patients with both kidney stones and consuming a high nephrolithiasis risk diet<sup>49</sup>). In a study examining the use of fasting as a treatment for high blood pressure in metabolic syndrome patients, *Hydrogenoanaerobacterium* was one of several taxa enriched in patients whose blood pressure was non-responsive to treatment<sup>50</sup>). However, the current state of the literature is largely comprised of simple correlational data with little in the way of mechanistic understanding of the impact of *Hydrogenoanaerobacterium* on host health.

Looking at the human clinical data, this bacterium seems to be viewed negatively. The decrease in this bacterium may be a positive effect.

### Genus *Parasutterella*

*Parasutterella* is an obligately anaerobic gram-negative genus and the most frequently reported taxa of *Betaproteobacteria* in the human gut, primarily represented by the species *P. excrementihominis*<sup>51</sup>). *P. excrementihominis* is asaccharolytic, primarily utilizing amino acids for energy metabolism. The bacterium plays a role in bile acid maintenance and cholesterol metabolism. It produces intermediate metabolites such as bile acid derivatives and succinate, which are utilized by other microbes in the gut.

*Parasutterella* shows numerous beneficial health effects as a symbiotic member of the gut microbiome. *Parasutterella* is significantly reduced by *C. difficile* carriage<sup>52</sup>), and is enriched in patients for whom fecal microbiota transplant successfully treated recurring *C. difficile* infection<sup>53</sup>). *Parasutterella* abundance is negatively correlated with clinical indicators of obesity and metabolic disorder, such as BMI and blood glucose level<sup>46, 54</sup>) and abundance is correlated with a reduction of serum lipids<sup>55</sup>). *Parasutterella* also appears to provide resistance against liver and kidney disease. In a mouse model of alcohol induced liver disease, microbiome transplants from alcoholic patients with and without hepatitis conferred susceptibility or resistance to liver damage, with *Parasutterella* the dominant genus of the hepatitis resistant group<sup>56</sup>). In patients with chronic kidney disease *Parasutterella* is significantly reduced compared to healthy controls, and its abundance is correlated with improved glomerular filtration rate and reduced levels of serum creatinine, blood urea nitrogen, and cystatin C<sup>57</sup>). During pregnancy, *P. excrementihominis* has neuroprotective effects on fetal development by increasing tryptophan metabolism in the gut and reducing kynurenine levels in maternal and fetal compartments<sup>58</sup>). *P. excrementihominis* is additionally enriched in healthy patients compared to those with pregnancy induced hypertension<sup>59</sup>).

Despite its otherwise healthy presence/effects in the gut, *Parasutterella* is associated with inflammation in IBS<sup>60</sup>), and is one of several enriched genera that are associated with autism spectrum disorder<sup>61</sup>) and depression<sup>62</sup>). However, current data is conflicting. Despite its association with inflammation in IBS and colitis, a recent study found an increase of *Parasutterella* and other *Lachnospiraceae* genera and their SCFA products, chiefly butyrate, accompanied amelioration of ulcerative colitis<sup>63</sup>). It is possible that there

was a compensatory increase in *Parasutterella* in response to the presence of disease.

There exists controversy about the role of these bacteria. Nevertheless, we consider that the increase in these bacteria is an overall positive.

### Genus *Ruminococcus*

*Ruminococcus* is a genus of anaerobic and gram-positive bacteria. *Ruminococcus* species abundance is positively correlated with SCFA production in the gut<sup>64</sup>; *R. albus* produces acetate from the breakdown of cellulose<sup>65</sup>, *R. bromii* ferments acetate from resistant starches<sup>66</sup>, and *R. gnavus* produces acetate, formate, lactate, and butyrate<sup>67,68</sup>. The health effects of *Ruminococcus* reported in the literature are mixed: the genus is enriched in type 2 diabetes<sup>69</sup>, however it appears to have a protective effect against type 1 diabetes via promotion of CD8+ Treg cell proliferation in murine models<sup>70</sup>. Ultimately, the outcomes of *Ruminococcus* abundance are heavily dependent on species level relationships, although the weight of the literature leans toward negative health associations.

*R. gnavus* is frequently reported in a pathogenic context, being significantly enriched in obesity<sup>71</sup>. Distinguishable strains of *R. gnavus* are linked with inflammatory bowel disease, and blooms of the bacteria are associated with increased severity of symptoms<sup>72,73</sup>. *R. gnavus* also produces a pro-inflammatory polysaccharide that induces cytokine upregulation, and dysbiosis of the species has been reported in association with increased inflammation in Crohn's disease<sup>74</sup>. Further, *R. gnavus* overabundance has been associated with the development of respiratory allergic disease in infants<sup>75</sup>, and mice fed *R. gnavus* develop histological signs of asthma<sup>76</sup>. Despite a generally negative influence on health, *R. gnavus* does have a positive effect in certain circumstances. It is reduced in infants with atopic dermatitis<sup>77</sup>, and *R. gnavus* oral administration alleviates atopic dermatitis severity in a mouse model of the disease by modulating immune response and increasing butyrate production<sup>78</sup>.

While *R. gnavus* is associated with Crohn's disease, other *Ruminococcus* species such as *R. albus*, *R. callidus*, and *R. bromii* are reduced in the disorder and are reported to be significantly more abundant in healthy controls<sup>79</sup>.

Recent literature has tended to characterize this bacterium as unfavorable to health, but this point requires further investigation.

### Phylum *Pseudomonadota* (Proteobacteria)

*Pseudomonadota* are a phylum of generally gram-negative bacteria, containing a highly diverse variety of bacterial species, including well-known genera *Escherichia*, *Salmonella*, and others. In contrast to the many obligate anaerobes found in the gut microbiome, there are numerous facultatively anaerobic species within *Pseudomonadota*. As such, *Pseudomonadota* are early colonizers of the gastrointestinal tract during infancy and facilitate the reshaping of the gut environment to better accommodate later colonization by depleting oxygen<sup>80</sup>.

*Pseudomonadota* tend to be present at relatively low abundance<sup>81</sup> compared to *Bacillota* (*Firmicutes*) and

*Bacteroidota*, however they are responsible for a significant amount of individual variation in functional genes of the gut microbiome<sup>82</sup>. While *Pseudomonadota* are commensal at low levels, when overabundant they can be a sign of intestinal dysbiosis<sup>81</sup> and are associated with disease states such as obesity<sup>83</sup>, diabetes<sup>84</sup>, Parkinson's<sup>85</sup>, and cognitive decline<sup>86,87</sup>.

Disorders involving elevated inflammation are also accompanied with *Pseudomonadota* dysbiosis. Respiratory diseases such as asthma involve dysbiosis of both the lung and gut microbiota, which is partially characterized by an overabundance of *Pseudomonadota* in both environments<sup>88,89</sup>. *Pseudomonadota* are significantly enriched while *Bacillota* are reduced in the gut of patients with irritable bowel syndrome<sup>90,91</sup>. Non-alcoholic Fatty Liver Disease is also associated with an elevated abundance of *Pseudomonadota* and microbial endotoxin production<sup>92,93</sup>.

While large blooms of the phylum are dysbiotic and contribute to the pathogenesis of a wide range of illnesses, at relatively low levels of abundance the *Pseudomonadota* are a core member of the gut microbiota play a vital role in the initial bacterial colonization of the intestines. A diverse taxon, *Pseudomonadota* and contains beneficial as well as pathogenic organisms.

This bacterium is thought to play an important role in maintaining the homeostasis of the intestinal microflora, so having an excess or deficiency is not desirable. If the deficient bacteria increase and return to a healthy state, this can be considered a positive effect. If the excess bacteria are reduced to a healthy state, this would also be favorable.

### Genus *Mediterraneibacter*

*Mediterraneibacter* is a genus of strictly anaerobic gram-positive bacteria, isolated from the fecal sample of an obese patient in France<sup>94</sup>. The type species is *M. massiliensis*, which produces acetate and isocaproate as the primary product of carbohydrate fermentation, while other species such as *M. butyricigenes*<sup>95</sup> produces butyrate. Other species include *M. hominis*<sup>96</sup>, and reclassifications of *M. faecis* and *M. glycyrrhizinilyticus* from *Ruminococcus* and *Clostridium* respectively<sup>94</sup>.

As a newly discovered genus, the role of *Mediterraneibacter* is still poorly understood. Early research indicates that it may be beneficial, as abundance of the genus is negatively correlated with severity of necrotic enteritis<sup>97</sup>. *M. faecis*, is a beneficial member of the gut microbiome, producing SCFAs<sup>98,99</sup> and thereby exerting antidiabetic<sup>100</sup> and anti-inflammatory effects<sup>101</sup>.

Based on these findings, this increase in bacteria can be interpreted as a positive effect.

### Genus *Monoglobus*

*Monoglobus pectinilyticus* is currently the only validly named species within the genus *Monoglobus*, isolated from a healthy human fecal sample in Korea<sup>102</sup>. *M. pectinilyticus* is a strictly anaerobic gram-positive bacterium characterized by its ability to ferment pectins, polysaccharides found in the walls of plant cells, which are abundant in citrus and other fruits. While unable to utilize glucose, the bacterium metabolizes fructose, producing the SCFAs acetate, formate, and lactate as metabolic byproducts.



*Monoglobus* appears to have a protective effect against liver disease. *Monoglobus* is significantly depleted in hepatitis B and hepatocellular carcinoma patients compared to healthy controls<sup>103</sup>. In children with biliary atresia, *Monoglobus* was significantly enriched in both patients who did not develop cholangitis after corrective surgery and in those whose jaundice cleared 6 months post-surgery<sup>104</sup>. *Monoglobus* is also correlated with Alzheimer's disease, being significantly reduced in AD patients compared to healthy controls: *Monoglobus* abundance was also negatively correlated with amyloid positivity<sup>105</sup> and blood triglyceride levels<sup>106</sup>. In patients with *C. difficile* infection, *Monoglobus* abundance was correlated with a reduction in infection severity and detectable *C. difficile* toxins<sup>107</sup>.

The relative abundance of *Monoglobus* has been reported to be elevated in rheumatoid arthritis, and was correlated with CD4<sup>+</sup> T cell counts and the levels of cytokines IL-2, IL-4, IL-10, TNF- $\alpha$ , and IFN- $\gamma$ <sup>108</sup>.

This increase in bacteria can be interpreted as a positive effect.

### Genus *Murimonas*

The Genus *Murimonas* is comprised of a single species, *Murimonas* intestini, isolated from the mouse intestine<sup>109</sup>. *M. intestine* is an obligate anaerobic gram-positive bacterium in the family *Lachnospiraceae* that produces the SCFA acetate. While not much is yet known of the role of this bacterium in the ecology of the gut microbiome, *Murimonas* is reported to be significantly depleted in obesity compared to normal weight controls<sup>110</sup>. Additionally, *Murimonas* may influence the gut-brain axis in depression, as its abundance is reportedly correlated with increased levels of Brain Derived Neurotrophic Factor<sup>111</sup> and is reduced by chronic unpredictable stress induced depression in mouse models of depression<sup>112</sup>.

*M. intestini* is an acetic acid-producing bacterium, and promotes BDNF production, which can be considered a positive effect.

### Comparison with dementia patients and those with poor sleep quality

In previous reports, a notable characteristic of those without dementia<sup>7</sup> and those with improved “sleep quality”<sup>5</sup> was the marked increase in *Bacteroidota* (Genus *Bacteroides*), with an increase in *Oscillospiraceae* (Clostridium cluster IV (Family *Ruminococcaceae*)) and *Lachnospiraceae* (Clostridium subcluster XIVa (Family *Lachnospiraceae*)) (Fig.1). Sleep quality and intestinal flora are known to have bidirectional effects mediated by the gut-brain axis. Improving “sleep quality” may help escape the dementia pattern of bacterial flora disorders and transition to a non-dementia pattern.

## Conclusion

Sleep quality and microbiota are closely linked by the gut-brain axis. In this report, we conducted a literature review of representative bacteria related to the gut-brain axis. It was suggested that improving sleep quality may increase SCFA-producing bacteria, which may help escape from dementia pattern dysbiosis and transition to a non-dementia pattern.

## Conflict of interest declaration

None.

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## Reference

- 1) Takabe W, Ogura M, Yagi M, et al. Effect on sleep quality of bedding with a high user rating in a post-marketing survey: A non-controlled open-label study. *Glycative Stress Res.* 2016; 3: 110-123.
- 2) Ogura M, Takabe W, Yagi M, et al. Effect of mats with “A Distinctive 4-Layer 3-Dimensional Structure” on sleep quality, anti-oxidative and immunological function. *Glycative Stress Res.* 2017; 4: 172-183.
- 3) Ogura M, Hattori A, Yagi M, et al. Effect of mats with “A Distinctive 4-Layer 3-Dimensional Structure” on sleep quality and nocturnal blood glucose: A crossover trial. *Glycative Stress Res.* 2019; 6: 49-63.
- 4) Ando M, Yagi M, Takabe W, et al. Effects of mats with “A Distinctive 4-Layer 3-Dimensional Structure” on sleep quality, skin function, and fatigue: A non-controlled open-label study. *Glycative Stress Res.* 2020; 7: 75-87, 2020.
- 5) Haasbroek K, Yagi M, Ando M, et al. Effects of mats with “A Distinctive 4-Layer 3-Dimensional Structure” on sleep quality and gut microbiota: A non-controlled open-label study. *Glycative Stress Res.* 2021; 8: 73-86.
- 6) Ogura M, Yagi M, Ando M, et al. Effects of mats with “A Distinctive 4-Layer 3-Dimensional Structure” on sleep quality and menopause: An open-label study. *Glycative Stress Res.* 2023; 10: 43-63.
- 7) Saji N, Niida S, Murotani K, et al. Analysis of the relationship between the gut microbiome and dementia: A cross-sectional study conducted in Japan. *Sci Rep.* 2019; 9: 1008.
- 8) Nakamura S, Ikeuchi T, Araki A, et al. Possibility for prevention of type 2 diabetes mellitus and dementia using three kinds of brown rice blends after high-pressure treatment. *Foods.* 2022; 11: 818.
- 9) Bhar S, Bose T, Dutta A, et al. A perspective on the benefits of consumption of parboiled rice over brown rice for glycaemic control. *Eur J Nutr.* 2022; 61: 615-624.
- 10) Benno Y, Endo K, Miyoshi H, et al. Effect of rice fiber on human fecal microflora. *Microbiol Immunol.* 1989; 33: 435-440.
- 11) Fukushima-Hirakawa A. Study on relationship between self-rated health and intestinal microbiota. *Int J Hum Cult Stud.* 2019; 29: 101-111. (in Japanese)
- 12) Akamine Y, Millman JF, Uema T, et al. Fermented brown rice beverage distinctively modulates the gut microbiota in Okinawans with metabolic syndrome: A randomized controlled trial. *Nutr Res.* 2022; 103: 68-81.
- 13) Zhao R, Fajardo J, Shen GX. Influence of brown or germinated brown rice supplementation on fecal short-chain fatty acids and microbiome in diet-induced insulin-resistant mice. *Microorganisms.* 2023; 11: 2629.
- 14) Flaiz M, Baur T, Brahner S, et al. *Caproicibacter* fermentans gen. nov., sp. nov., a new caproate-producing bacterium and emended description of the genus *Caproiciproducens*. *Int J Syst Evol Microbiol.* 2020; 70: 4269-4279.
- 15) Gilroy R, Ravi A, Getino M, et al. Extensive microbial diversity within the chicken gut microbiome revealed by metagenomics and culture. *PeerJ.* 2021; 9: e10941.
- 16) Tsubokawa M, Nishimura M, Mikami T, et al. Association of gut microbial genera with heart rate variability in the general Japanese population: The Iwaki Cross-Sectional Research Study. *Metabolites.* 2022; 12: 730.
- 17) Willing BP, Dicksved J, Halfvarson J, et al. A pyrosequencing study in twins shows that gastrointestinal microbial profiles vary with inflammatory bowel disease phenotypes. *Gastroenterology.* 2010; 139: 1844-1854.e1.
- 18) Zhang Z, Wang D, Qiao S, et al. Metabolic and microbial signatures in rat hepatocellular carcinoma treated with caffeic acid and chlorogenic acid. *Sci Rep.* 2017; 7(1): 4508.
- 19) Shen F, Zheng RD, Sun XQ, et al. Gut microbiota dysbiosis in patients with non-alcoholic fatty liver disease. *Hepatobiliary Pancreat Dis Int.* 2017; 16: 375-381.
- 20) Zhao X, Zhang Y, Guo R, et al. The Alteration in composition and function of gut microbiome in patients with type 2 diabetes. *J Diabetes Res.* 2020; 2020: 8842651.
- 21) Lu J, Zhang P, Hu R, et al. Gut microbiota characterization in Chinese patients with alopecia areata. *J Dermatol Sci.* 2021; 102: 109-115.
- 22) Jiang H, Ling Z, Zhang Y, et al. Altered fecal microbiota composition in patients with major depressive disorder. *Brain Behav Immun.* 2015; 48: 186-194.
- 23) Zheng P, Zeng B, Zhou C, et al. Gut microbiome remodeling induces depressive-like behaviors through a pathway mediated by the host's metabolism. *Mol Psychiatr.* 2016; 21: 786.
- 24) Zheng H, Liang H, Wang Y, et al. Altered gut microbiota composition associated with eczema in infants. *PLoS One.* 2016; 11: e0166026.
- 25) Rothenberg SE, Chen Q, Shen J, et al. Neurodevelopment correlates with gut microbiota in a cross-sectional analysis of children at 3 years of age in rural China. *Sci Rep.* 2021; 11: 7384.
- 26) Naito Y, Takagi T, Inoue R, et al. Gut microbiota differences in elderly subjects between rural city Kyotango and urban city Kyoto: An age-gender-matched study. *J Clin Biochem Nutr.* 2019; 65: 125-131.
- 27) Ueki A, Goto K, Kaku N, et al. *Aminipila butyrifica* gen. nov., sp. nov., a strictly anaerobic, arginine-decomposing bacterium isolated from a methanogenic reactor of cattle waste. *Int J Syst Evol Microbiol.* 2018; 68: 443-448.
- 28) Kim YB, Kim JY, Kim J, et al. *Aminipila terrae* sp. nov., a strictly anaerobic bacterium isolated from river sediment. *Arch Microbiol.* 2021; 203: 3163-3169.
- 29) Wei Z, Ma S, Chen R, et al. *Aminipila luticellarii* sp. nov., an anaerobic bacterium isolated from the pit mud of strong aromatic Chinese liquor, and emended description of the genus *Aminipila*. *Int J Syst Evol Microbiol.* 2021; 71(10).
- 30) Chen YJ, Hung WC, Chou YH, et al. Subgingival microbiome in rheumatoid arthritis Patients with periodontitis. *Int J Mol Sci.* 2022; 23: 9883.
- 31) Paz EA, Chua EG, Hassan SU, et al. Bacterial communities in the gastrointestinal tract segments of helminth-resistant and helminth-susceptible sheep. *Anim Microbiome.* 2022; 4: 23.
- 32) Sakamoto M. The Taxonomy of the Genus *Bacteroides* and related Taxa. *Journal of Intestinal Microbiology.* 2016; 30: 119-127. (in Japanese)
- 33) Sakamoto M, Takagaki A, Matsumoto K, et al. *Butyricimonas synergistica* gen. nov., sp. nov. and *Butyricimonas virosa* sp. nov., butyric acid-producing bacteria in the family 'Porphyromonadaceae' isolated from rat faeces. *Int J Syst Evol Microbiol.* 2009; 59: 1748-1753.

- 34) Sakamoto M, Tanaka Y, Benno Y, et al. *Butyricimonas faecihominis* sp. nov. and *Butyricimonas paravirosa* sp. nov., isolated from human faeces, and emended description of the genus *Butyricimonas*. *Int J Syst Evol Microbiol*. 2014; 64: 2992-2997.
- 35) Zhang W, Xu JH, Yu T, et al. Effects of berberine and metformin on intestinal inflammation and gut microbiome composition in db/db mice. *Biomed Pharmacother*. 2019; 118: 109131.
- 36) Yang S, Hu T, Liu H, et al. Akebia saponin D ameliorates metabolic syndrome (MetS) via remodeling gut microbiota and attenuating intestinal barrier injury. *Biomed Pharmacother*. 2021; 138: 111441.
- 37) Lee H, An J, Kim J, et al. A novel bacterium, *Butyricimonas virosa*, preventing HFD-induced diabetes and metabolic disorders in mice via GLP-1 receptor. *Front Microbiol*. 2022; 13: 858192.
- 38) Lv WQ, Lin X, Shen H, et al. Human gut microbiome impacts skeletal muscle mass via gut microbial synthesis of the short-chain fatty acid butyrate among healthy menopausal women. *J Cachexia Sarcopenia Muscle*. 2021; 12: 1860-1870.
- 39) Garcia-Mantrana I, Selma-Royo M, Alcantara C, et al. Shifts on gut microbiota associated to Mediterranean diet adherence and specific dietary intakes on general adult population. *Front Microbiol*. 2018; 9: 890.
- 40) Tian Y, Zuo L, Guo Q, et al. Potential role of fecal microbiota in patients with constipation. *Therap Adv Gastroenterol*. 2020; 13: 1756284820968423.
- 41) Jangi S, Gandhi R, Cox LM, et al. Alterations of the human gut microbiome in multiple sclerosis. *Nat Commun*. 2016; 7: 12015.
- 42) Schink M, Konturek PC, Tietz E, et al. Microbial patterns in patients with histamine intolerance. *J Physiol Pharmacol*. 2018; 69(4).
- 43) Dayama G, Priya S, Niccum DE, et al. Interactions between the gut microbiome and host gene regulation in cystic fibrosis. *Genome Med*. 2020; 12: 12.
- 44) Song L, Dong X. *Hydrogenoanaerobacterium saccharovorans* gen. nov., sp. nov., isolated from H<sub>2</sub>-producing UASB granules. *Int J Syst Evol Microbiol*. 2009; 59: 295-299.
- 45) Tung YC, Liang ZR, Yang MJ, et al. Oolong tea extract alleviates weight gain in high-fat diet-induced obese rats by regulating lipid metabolism and modulating gut microbiota. *Food Funct*. 2022; 13: 2846-2856.
- 46) Guo WL, Cao YJ, You SZ, et al. Ganoderic acids-rich ethanol extract from *Ganoderma lucidum* protects against alcoholic liver injury and modulates intestinal microbiota in mice with excessive alcohol intake. *Curr Res Food Sci*. 2022; 5: 515-530.
- 47) Jung MJ, Lee J, Shin NR, et al. Chronic repression of mTOR complex 2 induces changes in the gut microbiota of diet-induced obese mice. *Sci Rep*. 2016; 6: 30887.
- 48) Li F, Wang P, Chen Z, et al. Alteration of the fecal microbiota in North-Eastern Han Chinese population with sporadic Parkinson's disease. *Neurosci Lett*. 2019; 707: 134297.
- 49) Yuan C, Jin X, He Y, et al. Association of dietary patterns with gut microbiota in kidney stone and non-kidney stone individuals. *Urolithiasis*. 2022; 50: 389-399.
- 50) Maifeld A, Bartolomaeus H, Löber U, et al. Fasting alters the gut microbiome reducing blood pressure and body weight in metabolic syndrome patients. *Nat Commun*. 2021; 12: 1970.
- 51) Ju T, Kong JY, Stothard P, et al. Defining the role of *Parasutterella*, a previously uncharacterized member of the core gut microbiota. *ISME J*. 2019; 13: 1520-1534.
- 52) Zhang L, Dong D, Jiang C, et al. Insight into alteration of gut microbiota in *Clostridium difficile* infection and asymptomatic *C. difficile* colonization. *Anaerobe*. 2015; 34: 1-7.
- 53) Staley C, Kelly CR, Brandt LJ, et al. Complete microbiota engraftment is not essential for recovery from recurrent *Clostridium difficile* infection following fecal microbiota transplantation. *mBio*. 2016; 7: e01965-16.
- 54) Zeng Q, Li D, He Y, et al. Discrepant gut microbiota markers for the classification of obesity-related metabolic abnormalities. *Sci Rep*. 2019; 9: 13424.
- 55) Bush JR, Alfa MJ. Increasing levels of *Parasutterella* in the gut microbiome correlate with improving low-density lipoprotein levels in healthy adults consuming resistant potato starch during a randomised trial. *BMC Nutr*. 2020; 6: 72.
- 56) Llopis M, Cassard AM, Wrzosek L, et al. Intestinal microbiota contributes to individual susceptibility to alcoholic liver disease. *Gut*. 2016; 65: 830-839.
- 57) Li F, Wang M, Wang J, et al. Alterations to the gut microbiota and their correlation with inflammatory factors in chronic kidney disease. *Front Cell Infect Microbiol*. 2019; 9: 206.
- 58) Galley JD, Chen HJ, Antonson AM, et al. Prenatal stress-induced disruptions in microbial and host tryptophan metabolism and transport. *Behav Brain Res*. 2021; 414: 113471.
- 59) Lin H, Chen J, Ma S, et al. The association between gut microbiome and pregnancy-induced hypertension: A nested case-control study. *Nutrients*. 2022; 14: 4582.
- 60) Chen YJ, Wu H, Wu SD, et al. *Parasutterella*, in association with irritable bowel syndrome and intestinal chronic inflammation. *J Gastroenterol Hepatol*. 2018; 33: 1844-1852.
- 61) Pietrucci D, Teofani A, Milanesi M, et al. Machine learning data analysis highlights the role of *Parasutterella* and *Alloprevotella* in autism spectrum disorders. *Biomedicines*. 2022; 10: 2028.
- 62) Cheung SG, Goldenthal AR, Uhlemann AC, et al. Systematic review of gut microbiota and major depression. *Front Psychiatry*. 2019; 10: 34.
- 63) Liu X, Zhang Y, Li W, et al. Fucoidan ameliorated dextran sulfate sodium-induced ulcerative colitis by modulating gut microbiota and bile acid metabolism. *J Agric Food Chem*. 2022; 70: 14864-14876.
- 64) Yamamura R, Nakamura K, Kitada N, et al. Associations of gut microbiota, dietary intake, and serum short-chain fatty acids with fecal short-chain fatty acids. *Biosci Microbiota Food Health*. 2020; 39: 11-17.
- 65) Miller TL, Wolin MJ. Bioconversion of cellulose to acetate with pure cultures of *Ruminococcus albus* and a hydrogen-using acetogen. *Appl Environ Microbiol*. 1995; 61: 3832-3835.
- 66) Baxter NT, Schmidt AW, Venkataraman A, et al. Dynamics of human gut microbiota and short-chain fatty acids in response to dietary interventions with three fermentable fibers. *mBio*. 2019; 10: e02566-18.
- 67) Crost EH, Le Gall G, Laverde-Gomez JA, et al. Mechanistic insights into the cross-feeding of *Ruminococcus gnavus* and *Ruminococcus bromii* on host and dietary carbohydrates. *Front Microbiol*. 2018; 9: 2558.



- 68) Nilsen M, Madelen Saunders C, et al. Butyrate levels in the transition from an infant- to an adult-like gut microbiota correlate with bacterial networks associated with *Eubacterium rectale* and *Ruminococcus gnavus*. *Genes (Basel)*. 2020; 11: 1245.
- 69) Gurung M, Li Z, You H, et al. Role of gut microbiota in type 2 diabetes pathophysiology. *EBioMedicine*. 2020; 51: 102590.
- 70) Shimokawa C, Kato T, Takeuchi T, et al. CD8+ regulatory T cells are critical in prevention of autoimmune-mediated diabetes. *Nat Commun*. 2020; 11: 1922.
- 71) Jie Z, Yu X, Liu Y, et al. The baseline gut microbiota directs dieting-induced weight loss trajectories. *Gastroenterology*. 2021; 160: 2029-2042.e16.
- 72) Hall AB, Yassour M, Sauk J, et al. A novel *Ruminococcus gnavus* clade enriched in inflammatory bowel disease patients. *Genome Med*. 2017; 9: 103.
- 73) Henke MT, Brown EM, Cassilly CD, et al. Capsular polysaccharide correlates with immune response to the human gut microbe *Ruminococcus gnavus*. *Proc Natl Acad Sci USA*. 2021; 118: e2007595118.
- 74) Henke MT, Kenny DJ, Cassilly CD, et al. *Ruminococcus gnavus*, a member of the human gut microbiome associated with Crohn's disease, produces an inflammatory polysaccharide. *Proc Natl Acad Sci USA*. 2019; 116: 12672-12677.
- 75) Chua HH, Chou HC, Tung YL, et al. Intestinal dysbiosis featuring abundance of *Ruminococcus gnavus* associates with allergic diseases in infants. *Gastroenterology*. 2018; 154: 154-167.
- 76) De Filippis F, Paparo L, Nocerino R, et al. Specific gut microbiome signatures and the associated pro-inflammatory functions are linked to pediatric allergy and acquisition of immune tolerance. *Nat Commun*. 2021; 12: 5958.
- 77) Lee MJ, Kang MJ, Lee SY, et al. Perturbations of gut microbiome genes in infants with atopic dermatitis according to feeding type. *J Allergy Clin Immunol*. 2018; 141: 1310-1319.
- 78) Ahn JR, Lee SH, Kim B, et al. *Ruminococcus gnavus* ameliorates atopic dermatitis by enhancing Treg cell and metabolites in BALB/c mice. *Pediatr Allergy Immunol*. 2022; 33: e13678.
- 79) Kang S, Denman SE, Morrison M, et al. Dysbiosis of fecal microbiota in Crohn's disease patients as revealed by a custom phylogenetic microarray. *Inflamm Bowel Dis*. 2010; 16: 2034-2042.
- 80) Guaraldi F, Salvatori G. Effect of breast and formula feeding on gut microbiota shaping in newborns. *Front Cell Infect Microbiol*. 2012; 2: 94.
- 81) Shin NR, Whon TW, Bae JW. Proteobacteria: microbial signature of dysbiosis in gut microbiota. *Trends Biotechnol*. 2015; 33: 496-503.
- 82) Bradley PH, Pollard KS. Proteobacteria explain significant functional variability in the human gut microbiome. *Microbiome*. 2017; 5: 36.
- 83) Crovesy L, Masterson D, Rosado EL. Profile of the gut microbiota of adults with obesity: A systematic review. *Eur J Clin Nutr*. 2020; 74: 1251-1262.
- 84) Larsen N, Vogensen FK, van den Berg FW, et al. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. *PLoS One*. 2010; 5: e9085.
- 85) Sun MF, Zhu YL, Zhou ZL, et al. Neuroprotective effects of fecal microbiota transplantation on MPTP-induced Parkinson's disease mice: Gut microbiota, glial reaction and TLR4/TNF- $\alpha$  signaling pathway. *Brain Behav Immun*. 2018; 70: 48-60.
- 86) Nagpal R, Neth BJ, Wang S, et al. Modified Mediterranean-ketogenic diet modulates gut microbiome and short-chain fatty acids in association with Alzheimer's disease markers in subjects with mild cognitive impairment. *EBioMedicine*. 2019; 47: 529-542.
- 87) Shi H, Ge X, Ma X, et al. A fiber-deprived diet causes cognitive impairment and hippocampal microglia-mediated synaptic loss through the gut microbiota and metabolites. *Microbiome*. 2021; 9: 223.
- 88) Marsland BJ, Trompette A, Gollwitzer ES. The gut-lung axis in respiratory disease. *Ann Am Thorac Soc*. 2015; 12(Suppl 2): S150-S156.
- 89) Barcik W, Boutin RCT, Sokolowska M, et al. The Role of lung and gut microbiota in the pathology of asthma. *Immunity*. 2020; 52: 241-255.
- 90) Pittayanon R, Lau JT, Yuan Y, et al. Gut microbiota in patients with irritable bowel syndrome: A systematic review. *Gastroenterology*. 2019; 157: 97-108.
- 91) Vester-Andersen MK, Mirsepasi-Lauridsen HC, Prosberg MV, et al. Increased abundance of proteobacteria in aggressive Crohn's disease seven years after diagnosis. *Sci Rep*. 2019; 9: 13473.
- 92) Hoyles L, Fernández-Real JM, Federici M, et al. Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. *Nat Med*. 2018; 24: 1070-1080.
- 93) Vallianou N, Christodoulatos GS, Karampela I, et al. Understanding the role of the gut microbiome and microbial metabolites in non-alcoholic fatty liver disease: Current evidence and perspectives. *Biomolecules*. 2021; 12: 56.
- 94) Togo AH, Diop A, Bittar F, et al. Description of *Mediterraneibacter massiliensis*, gen. nov., sp. nov., a new genus isolated from the gut microbiota of an obese patient and reclassification of *Ruminococcus faecis*, *Ruminococcus lactaris*, *Ruminococcus torques*, *Ruminococcus gnavus* and *Clostridium glycyrrhizinilyticum* as *Mediterraneibacter faecis* comb. nov., *Mediterraneibacter lactaris* comb. nov., *Mediterraneibacter torques* comb. nov., *Mediterraneibacter gnavus* comb. nov. and *Mediterraneibacter glycyrrhizinilyticus* comb. nov. *Antonie Van Leeuwenhoek*. 2018; 111: 2107-2128.
- 95) Kim JS, Lee KC, Suh MK, et al. *Mediterraneibacter butyricigenes* sp. nov., a butyrate-producing bacterium isolated from human faeces. *J Microbiol*. 2019; 57: 38-44.
- 96) Liu C, Du MX, Abuduaini R, et al. Enlightening the taxonomy darkness of human gut microbiomes with a cultured biobank. *Microbiome*. 2021; 9: 119.
- 97) Yang Q, Liu J, Wang X, et al. Identification of an intestinal microbiota signature associated with the severity of necrotic enteritis. *Front Microbiol*. 2021; 12: 703693.
- 98) Jin M, Kalainy S, Baskota N, et al. Faecal microbiota from patients with cirrhosis has a low capacity to ferment non-digestible carbohydrates into short-chain fatty acids. *Liver Int*. 2019; 39: 1437-1447.
- 99) Ye X, Zhou L, Zhang Y, et al. Effect of host breeds on gut microbiome and serum metabolome in meat rabbits. *BMC Vet Res*. 2021; 17: 24.

- 100) Watanabe M, Sianoya A, Mishima R, et al. Gut microbiome status of urban and rural Filipino adults in relation to diet and metabolic disorders. *FEMS Microbiol Lett.* 2021; 368: fnab149.
- 101) Mahoney DE, Chalise P, Rahman F, et al. Influences of gastrointestinal microbiota dysbiosis on serum proinflammatory markers in epithelial ovarian cancer development and progression. *Cancers (Basel).* 2022; 14: 3022.
- 102) Kim CC, Kelly WJ, Patchett ML, et al. *Monoglobus pectinilyticus* gen. nov., sp. nov., a pectinolytic bacterium isolated from human faeces. *Int J Syst Evol Microbiol.* 2017; 67: 4992-4998.
- 103) Li R, Yi X, Yang J, et al. Gut microbiome signatures in the progression of hepatitis B virus-induced liver disease. *Front Microbiol.* 2022; 13: 916061.
- 104) Yang T, Yang S, Zhao J, et al. Comprehensive analysis of gut microbiota and fecal bile acid profiles in children with biliary atresia. *Front Cell Infect Microbiol.* 2022; 12: 914247.
- 105) Verhaar BJH, Hendriksen HMA, de Leeuw FA, et al. Gut microbiota composition is related to AD pathology. *Front Immunol.* 2022; 12: 794519.
- 106) Kaiyrylkyzy A, Kozhakhmetov S, Babenko D, et al. Study of gut microbiota alterations in Alzheimer's dementia patients from Kazakhstan. *Sci Rep.* 2022; 12: 15115.
- 107) Lesniak NA, Schubert AM, Flynn KJ, et al. The gut bacterial community potentiates *Clostridioides difficile* infection severity. *mBio.* 2022; 13: e0118322.
- 108) Wang Q, Zhang SX, Chang MJ, et al. Characteristics of the gut microbiome and its relationship with peripheral CD4+ T cell subpopulations and cytokines in rheumatoid arthritis. *Front Microbiol.* 2022; 13: 799602.
- 109) Kläring K, Just S, Lagkouvardos I, et al. *Murimonas intestini* gen. nov., sp. nov., an acetate-producing bacterium of the family Lachnospiraceae isolated from the mouse gut. *Int J Syst Evol Microbiol.* 2015; 65: 870-878.
- 110) Garcia-Ribera S, Amat-Bou M, Climent E, et al. Specific dietary components and gut microbiota composition are associated with obesity in children and adolescents with Prader-Willi Syndrome. *Nutrients.* 2020; 12: 1063.
- 111) Park M, Choi J, Lee HJ. Flavonoid-rich orange juice intake and altered gut microbiome in young adults with depressive symptom: A randomized controlled study. *Nutrients.* 2020; 12: 1815.
- 112) Liang XQ, Mai PY, Qin H, et al. Integrated 16S rRNA sequencing and metabolomics analysis to investigate the antidepressant role of Yang-Xin-Jie-Yu decoction on microbe-gut-metabolite in chronic unpredictable mild stress-induced depression rat model. *Front Pharmacol.* 2022; 13: 972351.