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Original article Effect of rice bran extract on *in vitro* advanced glycation end product formation

Masayuki Yagi¹⁾, Junko Naito¹⁾, Umenoi Hamada¹⁾, Sumie Okamura²⁾, Kazushi Yamamoto²⁾, Motonobu Matsumoto³⁾, Yoshikazu Yonei¹⁾

1) Anti-Aging Medical Research Center and Glycation Stress Research Center, Graduate School of Life and Medical Sciences, Doshisha University, Kyotanabe, Kyoto, Japan

2) New Material Business Development, Sunstar Co. Ltd., Takatsuki, Osaka, Japan

3) R&D, Health & Beauty Business Unit, Sunstar Co. Ltd., Takatsuki, Osaka, Japan

Abstract

Objectives: In recent years, attention has been paid to the effect of inhibiting advanced glycation end product (AGE) formation in the body for the purposes of anti-aging, health promotion, and lifestyle-disease prevention. In the present study, we evaluated the anti-glycation effects of rice bran extract and the potential of its use as an anti-glycation product.

Methods: The test product used was powdered rice bran provided by Sunbran Co., Ltd. (Tendo, Yamagata, Japan) and its hot water extract contains 37.83 mg/mL as solid content. Using an *in vitro* method with glucose and human serum albumin (HSA), we analyzed the inhibition of the formation of AGEs; specifically fluorescent AGEs, 3-deoxyglucosone (3DG), pentosidine (Pent), and N^{ε} -(carboxymethyl)lysine (CML) by rice bran extract utilizing fluorescence spectroscopy, high-performance liquid chromatography (HPLC), and enzyme linked immunosorbent assay (ELISA). We analyzed the test product's anti-glycation effects by comparing the half maximal inhibitory concentration (IC₅₀) against these glycated products with aminoguanidine, a known inhibitor of glycation.

Results: IC₅₀ of fluorescent AGEs (1.3 mg/mL) and 3DG (1.1 mg/mL) by rice bran (hot water extract) were 1.3 mg/mL and 1.1 mg/mL, respectively, 10 times higher than that of aminoguanidine; while IC₅₀ against CML (0.38 mg/mL) was 4 times higher for rice bran than that of aminoguanidine. IC₅₀ against Pent (3.8 mg/mL) was markedly lower for rice bran since aminoguanidine did not prevent Pent formation in this method.

Conclusion: These results suggest that rice bran extract inhibits glycation, although IC₅₀ was greater than that of aminoguanidine, except for Pent, and it may be useful for anti-glycation products.

KEY WORDS: rice bran, advanced glycation end products (AGEs), pentosidine, N^{*ε*}-(carboxymethyl)lysine (CML), 3-deoxyglucosone

Introduction

Rice bran is known to have a variety of components such as oil, protein, minerals and vitamin B group¹⁻⁶⁾. Rice bran oil characteristically contains a high ratio of oleic acid in fatty acid constitution and vitamin E, and it is reported to be resistant to oxidation by heating⁷⁾. However, no information is available regarding the effect of rice bran on glycative stress. The purpose of the present study is to elucidate the effect of the functional food material, a rice bran extract, on the formation of advanced glycation end products (AGEs) in an *in vitro* glycation model⁸⁾ between glucose and human serum albumin (HSA).

Method

Test product

The powdered rice bran "High-Bref [™]" (test product) was provided by Sunbran Co. Ltd. (Tendo, Yamagata, Japan). Hot water extract was prepared after adding 15 g test product to 150 mL distilled water followed by boiling for 1 hour at 100°C. The 75 mL extract solution was obtained after centrifugation at 3,000 rpm for 15 minutes and filtration of the supernatant by a Toyo filtration paper No.2 (Toyo Roshi Kaisha, Ltd., Bunkyo-ku, Tokyo, Japan); The extract solution was reserved in a freezer at -20°C until the experiment. The

Correspondence to: Professor Yoshikazu Yonei, MD, PhD

Anti-Aging Medical Research Center,

Graduate School of Life and Medical Sciences, Doshisha University

1-3 Tataramiyakodani, Kyotanabe-shi, Kyoto 610-0394 Japan

Phone/Fax: +81-77-465-6394 E-mail: yyonei@mail.doshisha.ac.jp

Co-authors: Yagi M, yagi@yonei-labo.com; Naito J, naito@yonei-labo.com;

Hamada U, hamada@yonei-labo.com; Okamura S, sumie.okamura@jp.sunstar.com;

Yamamoto K, kazushi.yamamoto@jp.sunstar.com;

Matsumoto M, motonobu.matsumoto@jp.sunstar.com

5 mL of hot water extract solution contained 37.83 mg/mL as solid content. The solution was diluted with distilled water and adjusted to the concentration of 12.61 mg/mL and 3.78 mg/mL. For the initial study using fluorescent AGE measurement, the ethanol extract was prepared after mixing 15 g test product and 150 mL of 80% ethanol and incubated at 37 °C for 15 hours with stirring (40 rpm/min); The reacted solution was centrifuged at 3,000 rpm for 15 minutes, then the supernatant was obtained. The 5 mL of ethanol extract solution contained 16.50 mg/mL as solid content. The solution was diluted with distilled water and adjusted to concentrations of 5.5 and 1.65 mg/mL. Aminoguanidine ⁹ (Wako Pure Chemical Industries Co., Ltd., Chuo-ku, Osaka) was used as a positive control.

Fluorescent AGE measurement

AGE-derived fluorescence was measured as reported previously using a glucose/HSA model 7). Briefly, 100 µL of various concentrations of the test product extract or aminoguanidine in aqueous solution were added to 500 μ L 0.1 mol/L phosphate buffered solution (PBS, pH 7.4), 100 µL distilled water, 200 µL 40 mg/mL HSA (Sigma Chemical Co., Ltd; St. Louis, MO, USA), and 100 µL 2.0 mol/L aqueous solution of glucose. Distilled water was then added to make up a total volume of 1.0 mL, and the material was incubated at 60°C for 40 hours (Solution A). Final concentrations were 8 mg/mL HSA and 0.2 mol/L glucose. At the same time, a solution including distilled water, added in lieu of aqueous glucose, was incubated as a blank for each reaction solution (Solution B). Samples prepared without the addition of the test product or aminoguanidine were incubated as positive controls (Solution C). And at the same time, a solution including distilled water, added in lieu of aqueous glucose, was incubated as a blank for each positive control (Solution D). Fluorescent AGEs were measured quantitatively in each sample reaction solution (A, B, C, D) to evaluate inhibitory activity for AGE formation. AGE-derived fluorescence was measured using an ARVO MX 1420 ARVO series Multilabel Counter (Perkin-Elmer Japan Corp.; Hodogaya-ku, Yokohama, Kanagawa, Japan) microplate reader at an excitation wavelength of 370 nm and a fluorescence wavelength of 440 nm. Calibration curves for inhibition of fluorescent AGE formation were constructed by adding individual samples to a reaction solution at three concentrations and calculating the inhibition of AGE formation after the reaction. Inhibition of AGE formation (%) was calculated using the following formula, and half maximal inhibitory concentration (IC50) values were then calculated to represent anti-glycation activity. Results were obtained as the average values of 3 measurements.

Formula: Inhibition of fluorescent AGE formation (%) = $(1-(A-B)/(C-D))\times 100$

3DG measurement

3-deoxyglucosone (3DG) was measured using the Tosoh high-performance liquid chromatography (HPLC) system (Tosoh Corporation, Minato-ku, Tokyo, Japan) as previously reported ^{10,11}. Samples were prepared from 100 μ L of test sample, aminoguanidine or distilled water added to 125 μ L of 20 μ g/mL 2,3-pentadione (Wako Pure Chemical Industries Ltd.), which was used as an internal standard, and 150 μ L of distilled water. The mixture was stirred, then 250 μ L of 6.0% perchloric acid (Wako Pure Chemical Industries) was added, stirred, and centrifuged at 12,000 rpm for 10 minutes; 800 μ L of the supernatant was added to 1,000 μ L of saturated sodium bicarbonate solution (Wako Pure Chemical Industries), stirred; 100 μ L of 2,3 diaminonaphthalene labeling reagent (Dojindo Laboratories, Kumamoto, Japan) was added, and the mixture was incubated for 24 hours at room temperature. The HPLC conditions were as follows; Column, YMC-Pack CN, 150 × 4.6 mm I.D column (YMC CO. Ltd., Shimogyo-ku, Kyoto, Japan); eluent, 50 mM phosphoric acid; acetonitrile: methanol = 70:17:13. The flow rate and detection wavelength were 1.0 mL/min and ultraviolet (UV) 268 nm.

Pentosidine measurement

Pentosidine (Pent) was measured by the enzyme-linked immunosorbent assay (ELISA) using a commercial kit "FSK Pentosidine" (Fushimi Pharmaceutical Co., Ltd., Marugame, Kagawa, Japan) as previously reported 10) and the manufacturer's instructions. After adding 50 μ L of serum or Pent standard to 20 µL pronase and 80 µL of Tris-HCl buffer, the mixture was incubated at 55°C for 90 minutes, and heated in boiling water for 15 minutes to inactive on the enzyme. These pretreated samples were added to each well and incubated at 37°C for 1 hour after washing. Then, 50 µL of anti-Pent antibody and Pent standard solution or pretreated sample were added to each well and incubated at 37°C for 1 hour after washing. A color development regent containing 0.5 mg/mL of 3,3'5,5'-tetramethylbenzidine (TMB) was added to each well. The reaction was stopped 10 minutes later by adding 100 μ L of TMB stop buffer. The absorbance was measured within 10 minutes at 450 nm and 630 nm. The standard curve was obtained by measuring standard Pent solutions.

CML measurement

 $N\varepsilon$ -(carboxymethyl)lysine (CML) was measured using a CircuLex NE-(carboxymethyl)lysine ELISA Kit (MBL Co. Ltd., Ina, Nagano, Japan)¹⁰⁾. Briefly, 30 µL of each reacted sample or CML standard were diluted with 90 μ L of sample/standard dilution buffer. Then, 120 µL of anti-CML adduct monoclonal antibody (clone name: MK-5A10) solution was added to each diluted sample, stirred, and 100 μ L of each mixture was dispensed into a well of an antigencoated microplate. The plates were incubated for 1 hour at room temperature, washed with 0.2% Tween-20; then 100 μ L of horse radish peroxidase-conjugated anti-mouse immunoglobulin G (IgG) polyclonal antibody solution was dispensed into each well, and further incubated for 1 hour, washed with washing buffer; then 100 μ L of tetra-methylbenzidine solution was added to each well, and the plate was wrapped in aluminum foil and incubated at room temperature for 10 minutes; 100 μ L of stop solution was added and the absorbance was measured at dual wavelengths of 450 nm/540 nm using a spectrophotometric microplate reader (SPECTRA MAX 190, Molecular Devices, Chuo-ku, Tokyo, Japan) within 30 minutes. The CML concentration in each sample was calculated from a standard curve of CML standards, and the IC 50 against CML by test samples was calculated.

Results

Fluorescent AGE measurement

The final concentrations of rice bran hot water extract were 0.378 mg/mL, 1.26 mg/mL and 3.78 mg/mL (*Fig.1*) and ethanol extract 0.165 mg/mL, 0.550 mg/mL, 1.65 mg/mL and 3.30 mg/mL (*Fig. 2*) in the *in vitro* reaction between glucose and HSA. The test product showed an inhibitory effect on fluorescent AGE formation in the glucose/HSA model; IC₅₀ was 1.3 mg/mL in hot water extract and 2.7 mg/mL in ethanol extract. The ethanol extract was less effective than the hot water extract. Aminoguanidine showed IC₅₀ of 0.095 mg/mL (*Fig. 3*), which was less than that of the test product.

3DG measurement

Percentage inhibition of rice bran hot water extract on 3DG formation is presented in *Fig. 4*. The extract dose-dependently inhibited 3DG formation in the glucose/HSA model; IC₅₀ was 1.1 mg/mL. Aminoguanidine also showed dose-dependent inhibition on 3DG formation; IC₅₀ was 0.12 mg/mL (*Fig. 5*).

Pent measurement

Percentage inhibition of rice bran hot water extract on Pent formation is presented in *Fig. 6*. The extract showed dose-dependent inhibition on Pent formation; IC₅₀ was 3.8 mg/mL in the glucose/HSA model. Aminoguanidine had no effect on Pent formation.

CML measurement

Percentage inhibition of rice bran hot water extract on CML formation is presented in *Fig.* 7. The extract dose-dependently inhibited CML formation in the glucose/HSA model; IC₅₀ was 0.38 mg/mL. Aminoguanidine showed dose-dependent inhibition on CML formation; IC₅₀ was 0.12 mg/ mL (*Fig. 8*).

Discussion

Rice bran extract contains a variety of active components which are reported to have an anti-oxidative effect ⁵), and an anti-inflammation effect ¹²) and improve glucose and lipid metabolism ¹³⁻¹⁵). The active components include polyphenols, such as protocatechuic acid, caffeic acid, ferulic acid, gentisic acid which have strong anti-oxidative capacity ⁵). Ferulic acid was the dominant phenolic acid in the rice bran samples.

This study added the anti-glycative effect of rice bran extract as an extra value associated to polyphenols. Some reports have shown the anti-glycation effect of ferulic acid ¹⁶⁻¹⁹. Caffeic acid oligomers reportedly prevent AGE formation by inhibiting the increase in 3-DG production ²⁰. Recently, rice bran was also found to contain quercetin ^{21,22} which is reported to show an anti-glycative effect ²³.

The results of IC₅₀ of the rice bran hot water extract showed a marked effect on Pent formation, one-fourth effect on CML formation and one-tenth effect on fluorescent AGE and 3DG formation, when compared with IC₅₀ of



Fig.1: Anti-glycation activity of rice bran hot water extract: Fluorescent AGE formation

Percent inhibition of fluorescent AGE formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC_{50}) = 1.3 mg/mL. AGE, advanced glycation end product; HSA, human serum albumin.



Fig.2: Anti-glycation activity of rice bran ethanol extract: Fluorescent AGE formation

Percent inhibition of fluorescent AGE formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC₅₀) = 2.7 mg/mL. AGE, advanced glycation end product; HSA, human serum albumin.



Fig.3: Anti-glycation activity of aminoguanidine: Fluorescent AGE formation

Percent inhibition of fluorescent AGE formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC $_{50}$) = 0.095 mg/mL. AGE, advanced glycation end product; HSA, human serum albumin.





Fig.4: Anti-glycation activity of rice bran hot water extract: 3DG formation

Percent inhibition of 3DG formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC50) = 1.1 mg/mL. 3DG, 3-deoxyglucosone; HSA, human serum albumin.



Fig.5: Anti-glycation activity of aminoguanidine: 3DG formation

Percent inhibition of 3DG formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC50) = 0.12 mg/ mL. 3DG, 3-deoxyglucosone; HSA, human serum albumin.



Fig. 6: Anti-glycation activity of rice bran hot water extract: Pent formation

Percent inhibition of Pent formation in the glucose/HSA reaction model. Half maximal inhibitory concentration $(IC_{50}) = 3.8$ mg/mL. Pent, pentosidine; HSA, human serum albumin.



Fig. 7: Anti-glycation activity of rice bran hot water extract: CML formation

Percent inhibition of 3DG formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC₅₀) = 0.38 mg/mL. CML, N^{ε} -(carboxymethyl)lysine; HSA, human serum albumin.



Fig.8: Anti-glycation activity of aminoguanidine: CML formation

Percent inhibition of CML formation in the glucose/HSA reaction model. Half maximal inhibitory concentration (IC₅₀) = 0.12 mg/mL. CML, N^{ε} -(carboxymethyl)lysine; HSA, human serum albumin.

aminoguanidine and rice-related samples ²⁴) as presented in *Table 1*. Rice bran (hot water extract) showed an equal IC₅₀ of rice (bran) and much less IC₅₀ than that of rice (unpolished and polished), almost ineffective, in the same experimental model ²⁴). The effect may be due to the contribution of rice bran ingredients, *i.e.*, ferulic acids. The mechanism by which these ferulic acids prevent AGE formation still remains unknown.

An intake of 150 - 300 mg/day of aminoguanidine, a glycation inhibitor, has been shown to have a progressioninhibiting effect on diabetic nephropathy²⁵⁾, and we previously calculated the daily intake of rice bran needed to obtain anti-glycation activity equivalent to that of aminoguanidine, resulting 4.2 g/day²⁴⁾. This amount of rice bran is not difficult to achieve to take orally.

In the present study, the rice bran extract inhibited CML formation more strongly than others, *i.e.*, fluorescent AGEs, 3DG, or pentosidine. In the AGE formation process, oxidative reactions are involved, especially with CML formation²⁶⁾. The reason why the effect was marked on CML formation may be explained by the anti-oxidation effects of polyphenols which are rich in rice bran in addition to its anti-glycation effect.

Conclusion

Rice bran hot water extract was confirmed to prevent AGE formation by inhibiting 3DG production in the *in vitro* glucose/HSA model. The mechanism remains unknown. Although IC₅₀ of the test product was higher than that of aminoguanidine, except for Pent, it may be useful as an anti-glycation product.

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Conflicts of interest statement

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No.	Sample name	IC 50 (mg/mL)			
		Fluorescent AGEs	3DG	Pent	CML
1	Rice bran (hot water extract)	1.3	1.1	3.8	0.38
2	Aminoguanidine	0.095	0.12	NE	0.12
3	Rice (unpolished)*	203.557			
4	Rice (bran)*	1.409			
5	Rice (polished)*	NE			

Table 1. Comparison of anti-glycation activity in the in vitro glucose / HSA model

 $IC_{50}, half maximal inhibitory concentration; HSA, human serum albumin; AGEs, anti-glycation activity; 3DG, 3-deoxyglucosone; Pent, pentosidine; CML, N&-(carboxymethyl)lysine; NE, no effect.$

*Data of rice-related samples (unpolished, bran, and polished) are from reference 24.

References

- Morita K, Hirakawa H, Matsueda T, et al. Stimulating effect of dietary fiber on fecal excretion of polychlorinated dibenzofurans (PCDF) and polychlorinated dibenzo-pdioxins (PCDD) in rats. Fukuoka Igaku Zasshi. 1993; 84: 273-281.(in Japanese)
- 2) Sugano M, Tsuji E. Rice bran oil and cholesterol metabolism. J Nutr. 1997; 127: 521S-524S.
- Sugano M, Koba K, Tsuji E. Health benefits of rice bran oil. Anticancer Res. 1999; 19: 3651-3657.
- Koba K, Liu JW, Bobik E, et al. Cholesterol supplementation attenuates the hypocholesterolemic effect of rice bran oil in rats. J Nutr Sci Vitaminol (Tokyo). 2000; 46: 58-64.
- Okai Y, Higashi-Okai K. Radical-scavenging activity of hot water extract of Japanese rice bran: Association with phenolic acids. J UOEH. 2006; 28: 1-12.
- 6) Kawakami Y, Tsuzuki T, Nakagawa K, et al. Distribution of tocotrienols in rats fed a rice bran tocotrienol concentrate. Biosci Biotechnol Biochem. 2007; 71: 464-471.
- Friedman M. Rice brans, rice bran oils, and rice hulls: composition, food and industrial uses, and bioactivities in humans, animals, and cells. J Agric Food Chem. 2013; 61: 10626-10641.
- Hori M, Yagi M, Nomoto K, et al. Experimental models for advanced glycation end product formation using albumin, collagen, elastin, keratin and proteoglycan. Anti-Aging Medicine. 2012; 9: 125-134.
- Nilsson BO. Biological effects of aminoguanidine: an update. Inflamm Res. 1999; 48: 509-515.
- 10) Yonei Y, Yagi M, Hibino S, et al. Herbal extracts inhibit Maillard reaction, and reduce chronic diabetic complications risk in streptozotocin-induced diabetic rats. Anti-Aging Medicine. 2008; 5: 93-98.
- Kitano H, Yagi M, Nomoto K, et al. Research on the inhibitory effect of edible purple chrysanthemum on generation of advanced glycation end products (AGEs). New Food Industry. 2011; 53: 1-10. (in Japanese)
- 12) Sakai S, Murata T, Tsubosaka Y, et al. γ -Oryzanol reduces adhesion molecule expression in vascular endothelial cells via suppression of nuclear factor- κ B activation. J Agric Food Chem. 2012; 60: 3367-3372.
- 13) Hongu N, Kitts DD, Zawistowski J, et al. Pigmented rice bran and plant sterol combination reduces serum lipids in overweight and obese adults. J Am Coll Nutr. 2014; 33: 231-238.
- 14) Chandrashekar P, Kumar PK, Ramesh HP, et al. Hypolipidemic effect of oryzanol concentrate and low temperature extracted crude rice bran oil in experimental male wistar rats. J Food Sci Technol. 2014; 51: 1278-1285.
- Berraaouan A, Abid S, Bnouham M. Antidiabetic oils. Curr Diabetes Rev. 2013; 9: 499-505.
- 16) Srey C, Hull GL, Connolly L, et al. Effect of inhibitor compounds on N^ε-(carboxymethyl)lysine (CML) and N^ε-(carboxyethyl)lysine (CEL) formation in model foods. J Agric Food Chem. 2010; 58: 12036-12041.
- 17) Silván JM, Assar SH, Srey C, Dolores Del Castillo M, Ames JM. Control of the Maillard reaction by ferulic acid. Food Chem. 2011; 128: 208-213.

- 18) Sompong W, Meeprom A, Cheng H, et al. A comparative study of ferulic acid on different monosaccharide-mediated protein glycation and oxidative damage in bovine serum albumin. Molecules. 2013; 18: 13886-13903.
- 19) Maruf AA, Lip H, Wong H, et al. Protective effects of ferulic acid and related polyphenols against glyoxal- or methylglyoxal-induced cytotoxicity and oxidative stress in isolated rat hepatocytes. Chem Biol Interact. 2014 Nov 18. pii: S0009-2797(14)00356-1. doi: 10.1016/ j.cbi.2014.11.007. [Epub ahead of print]
- 20) Sasaki K, Chiba S, Yoshizaki F. Effect of natural flavonoids, stilbenes and caffeic acid oligomers on protein glycation. Biomed Rep. 2014; 2: 628-632.
- 21) Moon HI, Lee JH, Lee YC, et al. Inhibitory effects of isolated compounds from black coloured rice bran on the complement classical pathway. Phytother Res. 2011; 25: 1418-1420.
- 22) Sriseadka T, Wongpornchai S, Rayanakorn M. Quantification of flavonoids in black rice by liquid chromatography-negative electrospray ionization tandem mass spectrometry. J Agric Food Chem. 2012; 60: 11723-11732.
- 23) Johari A, Moosavi-Movahedi A, Amanlou M. Computational investigation of inhibitory mechanism of flavonoids as bovine serum albumin anti-glycation agents. Daru. 2014 Dec 11;22(1):79. [Epub ahead of print]
- 24) Ishioka Y, Yagi M, Ogura M, et al. Antiglycation effect of various vegetables: Inhibition of advanced glycation end product formation in glucose and human serum albumin reaction system. Glycative Stress Research 2015; 2: 22-00.
- 25) Bolton WK, Cattran DC, Williams ME, et al. Randomized trial of an inhibitor of formation of advanced glycation end products in diabetic nephropathy. Am J Nephrol. 2004; 24: 32-40.
- 26) Fu MX, Requena JR, Jenkins AJ, et al. The advanced glycation end product, N^ε-(carboxymethyl)lysine, is a product of both lipid peroxidation and glycoxidation reactions. J Biol Chem. 1996; 271: 9982-9986.